

Novel sulindac derivatives for colorectal cancer chemoprevention that target cGMP phosphodiesterases to suppress Wnt/ β -catenin transcriptional activity

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ABSTRACT

Approximately 28 million individuals in the United States face the risk of developing precancerous colonic adenomas (polyps) and potentially progressing to colorectal cancer (CRC). While a promising strategy for CRC prevention involves pharmacological intervention, such as cancer chemoprevention or interception, currently, there are no FDA-approved drugs capable of preventing the formation or progression of adenomas to adenocarcinoma. Numerous clinical, epidemiological, and preclinical studies have offered compelling evidence supporting the efficacy of nonsteroidal anti-inflammatory drugs (NSAIDs) in CRC chemoprevention. However, the prolonged use of NSAIDs is not FDA-approved due to potential life-threatening toxicities resulting from cyclooxygenase (COX) inhibition and the depletion of physiological prostaglandins. Despite indications that the COX inhibitory activity of NSAIDs may not be essential for their antineoplastic effects, the absence of a well-defined target impeded the development of derivatives that do not inhibit COX. Earlier research suggests that the inhibition of cyclic guanosine monophosphate phosphodiesterase (cGMP PDE) may be responsible, at least in part, for the antineoplastic activity of the NSAID sulindac. This could potentially offer a novel target for CRC chemoprevention. To identify the cGMP PDE isozyme(s) contributing to the antineoplastic activity of sulindac, we synthesized a chemically diverse library of over 1500 compounds, all sharing the indene scaffold of sulindac. Subsequently, we screened these compounds for their impact on cancer cell growth and PDE inhibitory activity. From this screening, a series of lead compounds emerged. These compounds lacked COX-1 and COX-2 inhibitory activity, surpassing sulindac in potency to inhibit CRC cell growth. Importantly, they demonstrated greater selectivity by not affecting normal cell growth. Through chemical optimization, we identified several development candidates that selectively inhibit PDE5 and/or PDE10. These compounds activate cGMP/PKG signaling, suppressing Wnt/ β -catenin transcription. This action counters the growth advantages resulting from APC or CTNNB1 mutations, which are responsible for most human CRCs. This review delves into the scientific literature supporting PDE5 and/or PDE10 as potential targets for CRC chemoprevention or interception. Our findings suggest a promising avenue for developing drugs that may effectively intervene in the progression of colorectal cancer, offering hope for improved preventive strategies in the future.

KEYWORDS

cGMP; chemoprevention; colorectal cancer; cyclooxygenase; NSAIDs; phosphodiesterase; PKG; sulindac; Wnt; β -catenin

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1. Introduction

NSAIDs are among the most frequently prescribed drugs worldwide for the treatment of pain, fever, or inflammation. Epidemiologic studies have demonstrated that long-term use of NSAIDs, including aspirin and other non-prescription NSAIDs, can significantly reduce the risk of developing CRC and other cancers [1-6]. In addition, randomized clinical trials involving patients with a rare hereditary precancerous condition, familial adenomatous polyposis (FAP), demonstrated that two prescription NSAIDs, sulindac, a non-selective cyclooxygenase (COX) inhibitor [7-11], and celecoxib, a selective cyclooxygenase-2 (COX-2) inhibitor [12] could suppress the formation and/or cause the regression of precancerous adenomatous polyps. The cancer chemopreventive activity of NSAIDs and COX-2 inhibitors is also supported by multiple lines of evidence from preclinical studies using various experimental models.

Although NSAIDs have shown promising cancer chemopreventive activity, none are FDA-approved for long-term use because of potentially fatal toxicities associated with the inhibition of COX-1 and/or COX-2 enzymes that catalyze the conversion of arachidonic acid to prostaglandins and other eicosanoids. Prostaglandins are well known to have an essential role in normal physiological processes, including gastric protection and kidney function, but are also implicated in colon tumorigenesis by altering cell adhesion, impeding apoptosis, and promoting angiogenesis [13, 14]. In addition, prostaglandin levels are elevated in adenomas and adenocarcinomas [15] due to COX-2 overexpression, which supports the rationale to target COX-2 for CRC chemoprevention. The *Apc* Δ 716 mouse model of CRC provides further evidence for the role of COX-2 in the development of intestinal adenomas because its adenomatous phenotype can be partially suppressed by COX-2 knockdown [16].

While the cancer chemopreventive activity of NSAIDs and COX-2 inhibitors is commonly attributed to COX-2 inhibition, multiple lines of evidence suggest that an off-target effect may partially or fully account for their antineoplastic activity. For example, dosages exceeding those required for COX inhibition are required to inhibit CRC cell growth. Perhaps the most compelling evidence for a COX-independent mechanism comes from studies reporting that the non-COX inhibitory sulfone metabolite of sulindac has cancer chemopreventive activity in multiple rodent models of chemical-induced carcinogenesis [10, 17-22]. Hence, in addition to COX-1 and COX-2, multiple other targets, along with accompanying signaling pathways, have been suggested to be responsible for the antineoplastic activity of NSAIDs and COX-2 inhibitors including cGMP PDE, RAS, PPAR δ , and NF- κ B to name a few [23-29]. This review focuses on cGMP PDE isozymes as novel targets for CRC, which, as illustrated in [Figure 1](#) can be inhibited by sulindac, other NSAIDs, sulindac sulfone, and certain non-COX inhibitory derivatives to induce cGMP/PKG signaling. Activated PKG can phosphorylate β -catenin, triggering ubiquitination and proteasomal degradation of β -catenin to block TCF/LEF transcriptional activity, thereby countering the stabilization of β -catenin resulting from mutations in APC or β -catenin responsible for the majority of CRC cases [19, 30-33].

2. FAP and NSAIDs

FAP is an autosomal dominant syndrome, which is the second most common inherited CRC syndrome with an incidence of 1 in 10,000 [34] characterized by a high prevalence of adenomas or “polyps”. Clinical management of this disease involves frequent colonoscopies that are expensive, uncomfortable, and potentially fatal because of accidental perforation of the gastrointestinal tract. Surgical resection of the colon is often performed to prevent malignant progression in FAP patients at an early age. Pharmacological intervention has the potential to minimize the frequency of colonoscopies and delay surgical intervention. Medications effective for preventing colon adenomas would also be expected to benefit individuals with sporadic adenomatous polyposis and be particularly valuable in developing countries where screening by routine colonoscopy is not easily accessible. Based on experimental and clinical studies, the reduction of polyp burden by a cancer chemopreventive drug would also

reduce the risk of developing CRC in these high-risk populations.

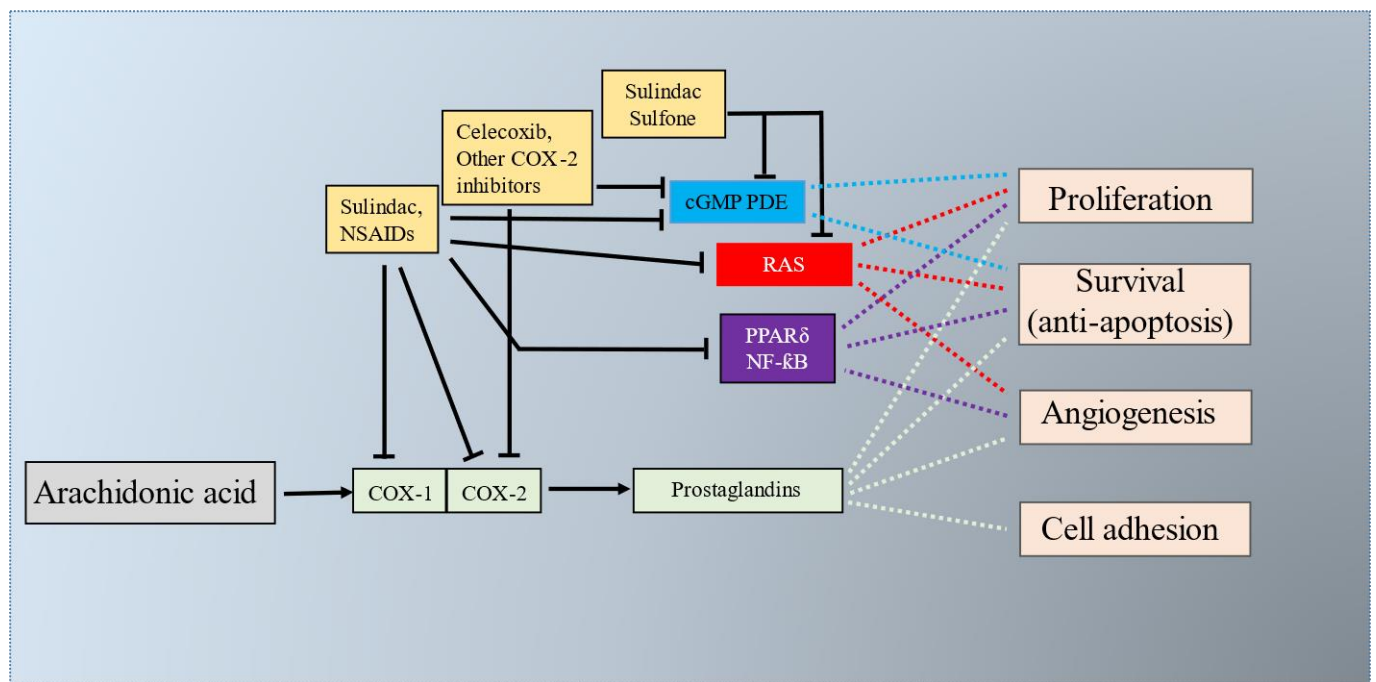


Figure 1. Potential COX-dependent and independent targets responsible for the cancer chemopreventive activity of NSAIDs, COX-2 inhibitors, sulindac sulfone, and derivatives. Blocking (indicated by $-$ symbol) of molecular targets results in modulation (indicated by dashed lines $---$) of the pathologic and physiologic processes.

FAP is attributed to loss-of-function mutations in adenomatous polyposis coli (APC), a tumor suppressor gene, which plays a pivotal role in regulating β -catenin transcriptional activity. The stability of cytoplasmic β -catenin is controlled by a molecular complex commonly referred to as the “destruction complex” comprised of APC, axin, glycogen synthase kinase 3 (GSK3), and casein kinase 1 (CK1) (Figure 2) [35]. The complex serves to regulate cytoplasmic levels of β -catenin by ubiquitin-dependent proteasomal degradation. Loss of APC tumor suppressor function resulting from APC gene mutations stabilizes β -catenin and promotes translocation to the nucleus to activate TCF/LEF mediated transcription of multiple growth-promoting and survival genes resulting in dysplasia and the formation of adenomas [36, 37]. Additional mutations in genes encoding for p53 and K-RAS and other oncoproteins arise that provide proliferation and survival advantages to precancerous cells, which have the potential to advance from dysplasia to adenomas to adenocarcinoma [38].

Randomized controlled clinical trials have shown that sulindac can significantly reduce the number and size of colon adenomas in FAP patients [8]. The efficacy of celecoxib, a selective COX-2 inhibitor, was also demonstrated in FAP patients [12, 39, 40], although at dosages appreciably higher than sulindac. The FDA previously approved celecoxib for the treatment of FAP; however, the approval was later withdrawn because of the increased risk of mortality from cardiovascular toxicity associated with COX-2 inhibitors and other NSAIDs [41].

2.1. COX-independent antitumor activity of NSAIDs

COX-1 and COX-2 isoforms have similar enzymatic activity but differ in how their expression is regulated and in their sensitivity to inhibitors [42]. COX-1 is constitutively expressed in a wide range of tissues, while the expression of COX-2 is generally low in normal tissues but induced by cytokines or stress in various tissues. COX-1 maintains prostaglandin levels essential for various physiological processes, including gastric protection and kidney function. As such, the depletion of prostaglandins by non-selective COX inhibitors can cause ulceration of the gastric mucosa, leading to gastrointestinal perforation or vasoconstriction at the glomerular level to reduce renal plasma

flow, resulting in kidney damage. By contrast, COX-2 is an inducible enzyme in which its expression is regulated at the transcriptional level [43]. COX-2 expression can be induced by cytokines under inflammatory conditions and is elevated in cancer cells to promote tumor growth [44], inhibit apoptosis [45], and induce angiogenesis [13, 46]. Consequently, COX-2 was assumed to be an attractive target for inflammation and cancer prevention whereby COX-2 inhibitors would have selective anti-inflammatory and antitumor activities. However, clinical trials of the COX-2 inhibitor rofecoxib (Vioxx) revealed an unexpected risk of cardiovascular toxicity that led to its withdrawal from the market and increased scrutiny for the long-term use of other COX-2 inhibitors, as well as non-selective NSAIDs, for cancer chemoprevention [41, 47, 48].

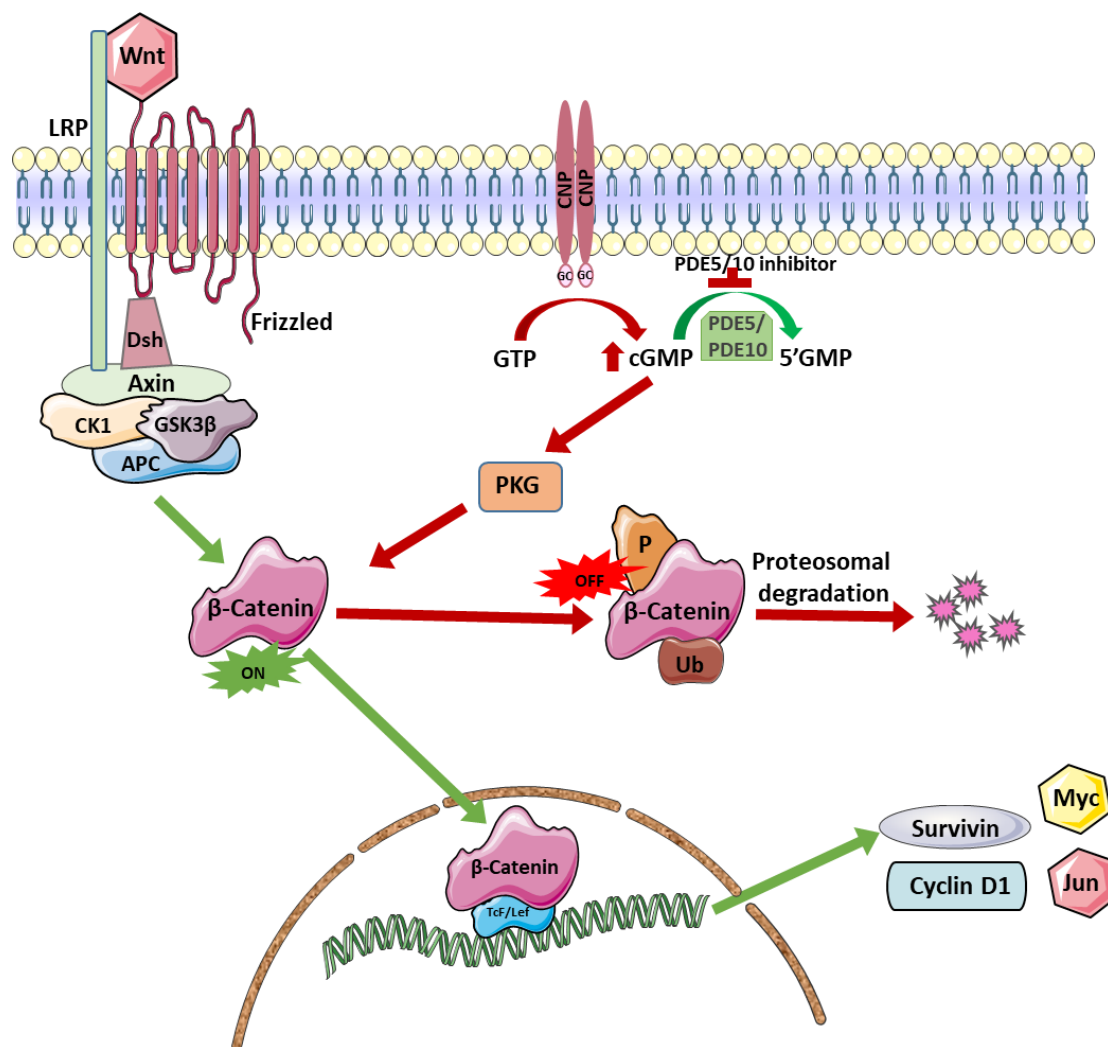


Figure 2. Loss of function mutations in the tumor suppressor gene, *APC*, or gain-in-function mutations in the oncogene, *CTNNB1* encode for proteins that result in the aberrant stabilization of β -catenin in the cytoplasm and translocation to the nucleus to activate TCF/LEF mediated transcription of proteins essential for colon tumorigenesis. PDE5 and PDE10 are overexpressed during the early stages of colon tumorigenesis and function to regulate intracellular cGMP levels, whereby low levels appear essential for the proliferation and survival of neoplastic cells. PDE5 and PDE10 inhibitors increase intracellular cGMP levels to activate PKG that can phosphorylate β -catenin on amino acid residues that induce ubiquitination and proteasomal degradation of β -catenin. Targeting PDE5 and/or PDE10 with non-COX inhibitory sulindac derivatives provides a strategy to develop drugs for CRC chemoprevention given that the proliferation and survival of neoplastic cells is dependent on TCF/LEF transcription factors that encode for oncogenic proteins such as cyclin D, survivin, Jun and Myc.

Elder and colleagues originally suggested evidence for a COX-2 independent mechanism responsible for the antineoplastic activity of COX-2 selective inhibitors, reporting that their apoptosis-inducing activity is unrelated to COX-2 expression levels [49]. Other investigators showed that supplementation with prostaglandins does not reverse the growth-inhibitory activity of NSAIDs [50, 51]. In addition, the rank order of potency among a series of chemically distinct NSAIDs and COX-2 inhibitors to inhibit CRC cell growth and COX-2 does not match: and that inhibition of CRC cell growth typically requires appreciably higher dosages than those required to inhibit prostaglandin synthesis [52-54].

Other investigators reported that sulindac sulfone, a non-COX inhibitory metabolite of sulindac, inhibits tumor formation in a variety of preclinical rodent models of chemical-induced carcinogenesis, providing additional evidence for a COX-independent mechanism of action [10, 17-22]. Because of reduced likelihood for gastrointestinal and renal toxicity resulting from the depletion of prostaglandins, sulindac sulfone (exisulind) was advanced to clinical trials in FAP patients by Cell Pathways Inc. Exisulind demonstrated promising inhibition or regression of colon adenomas in early clinical trials [55], but did not receive FDA approval because of hepatotoxicity observed in a Phase III registration trial in addition to a low response rate. A 12-month multicenter randomized double-blind placebo-controlled phase 3 dose-response study involving subjects with sporadic polyps also reported modest efficacy of exisulind by the decrease in median polyp size which was significantly greater in patients who received exisulind compared with those who received placebo. [56]. The limited efficacy of exisulind as observed in clinical trials, was likely attributable to the low potency [57]. A more potent analog of exisulind, CP-461, was later developed by Cell Pathways Inc., but poor oral bioavailability halted its clinical development [58]. Despite these failures, mechanistic studies of exisulind, CP-461, and other highly potent derivatives (e.g., CP-248) revealed a strong association between their growth inhibitory activity in CRC cells and the inhibition of cGMP PDE activity [32], although it was unclear which PDE isozyme(s) were required for colon cancer growth that might provide a vulnerability to selectively target for CRC chemoprevention.

2.2. Sulindac derivatives lacking COX-inhibitory activities

An initial attempt to develop novel sulindac derivatives involved molecular modeling using crystal structures of COX-1 and COX-2 to interfere with COX-1 and COX-2 inhibition. These studies revealed an essential role of the carboxylic acid moiety of sulindac sulfide to bind both COX-1 and COX-2, which guided the design of sulindac sulfide amide (SSA) that was appreciably more potent than sulindac sulfide to inhibit CRC growth [59]. The positively charged N, N-dimethyl ethyl ammonium moiety in SSA that replaced the negatively charged carboxylic acid moiety in sulindac was sufficient to eliminate both COX-1 and COX-2 inhibitory activity while improving potency to inhibit CRC cell growth. SSA was well tolerated and demonstrated *in vivo* antitumor activity in rodent models of colon, breast, and prostate cancer, although with modest potency that was attributed to limited oral bioavailability [59-61].

Further chemical optimization with adaptations and substitutions to different parts of the indene scaffold was performed to improve potency and oral bioavailability. This effort resulted in a custom library of over 1500 indenenes that were screened for growth inhibition using human CRC cell lines HT29, HCT116, and SW480, while NCM-460 normal colon mucosa cells were used to assess cancer cell selectivity. Testing a wide range of concentrations allowed the measurement of accurate IC₅₀ values to define specific chemical properties necessary for potency and cancer cell selectivity. Most of the compounds in the library were significantly more potent than sulindac sulfide with low micromolar IC₅₀ values to inhibit growth but with poorly defined structure-activity relationships (SAR). However, a discrete set of compounds with well-defined SAR and high potency and selectivity for CRC cell growth inhibition were identified by counter-screening in enzymatic assays that evaluated cGMP PDE inhibitory activity using tumor cell lysates. Compounds of interest were further screened using recombinant PDE isozymes to assess isozyme

selectivity. This chemical-biology approach aimed to identify specific PDE isozymes that represent potential targets for CRC chemoprevention.

3. Targeting PDE5 and PDE10 for CRC chemoprevention

3.1. Role of PDE5 in regulating CRC growth

Previous studies suggested that cGMP PDE inhibition is closely associated with the activity of sulindac sulfide and sulfone to inhibit CRC cell growth [32, 53, 62]. In these experiments, cancer cell growth measured using multiple cancer cell lines, while cGMP PDE enzymatic activity was measured using recombinant PDE isozymes. This methodology and other experimental procedures revealed a novel mechanism by which PDE5 inhibition can selectively inhibit CRC cell growth. As illustrated in Figure 2, guanylyl cyclases (GCs), which are widely distributed and activated in response to nitric oxide or natriuretic peptides from the tumor microenvironment, regulate physiologically cGMP levels in cells. The effects of endogenous GC activators can be augmented by cGMP PDE inhibition, thus increasing intracellular cGMP levels and PKG activation. The oncoprotein β -catenin appears to be an essential substrate of PKG. PKG-mediated phosphorylation of β -catenin can cause ubiquitination and proteasomal degradation of the "oncogenic" (nonphosphorylated, stable) pool of β -catenin and inhibit Wnt-induced Tcf/Lef transcription of genes involved in cancer cell survival and proliferation, including those encoding survivin and cyclin D. In addition to providing insight to the mechanism of growth suppression from PDE5 inhibition, these studies also revealed that cAMP PDE inhibitors/PKA activators do not appreciably affect CRC cell growth, while inhibitors of cGMP PDE isozymes could suppress CRC cell growth. PDE5 was implicated in the growth inhibitory activity of sulindac sulfide and sulfone based on observations showing overexpression in CRC cells compared to cells from normal colon mucosa [53, 63]. Sulindac sulfide also directly inhibited recombinant PDE5 at concentrations comparable to those that inhibit CRC cell growth [53, 64]. Furthermore, a novel sulindac derivative, sulindac benzylamine (SBA) was identified which lacked significant COX-1 and COX-2 inhibitory action but inhibited PDE5 and CRC cell growth, and, was able to suppress β -catenin transcriptional activity [63].

Consistent with the growth and PDE5 inhibitory activity of sulindac sulfide and SBA, genetic silencing of PDE5 in CRC cells was found to suppress CRC cell growth by activating cGMP/PKG signaling and inhibiting β -catenin transcriptional activity [63, 65]. In support of these observations implicating PDE5 as a novel anticancer target, other investigators reported that the highly specific PDE5 inhibitor, sildenafil, can inhibit CRC cell growth and tumor formation *in vitro* and *in vivo*, respectively, although at higher concentrations than those required to inhibit recombinant PDE5 [66]. The high dosages of sildenafil required to inhibit CRC cell growth may be attributed to efflux and low intracellular levels, given that PDE5 inhibitors can act as substrates for specific ABC transport proteins that may be overexpressed in cancer cells [67].

In support of the role of PDE5 in CRC, sildenafil was reported to inhibit inflammation-induced colon carcinogenesis in animal models by two independent groups using pharmacologically relevant concentrations [68, 69]. Recent epidemiological studies showing that PDE5 inhibitors can reduce the risk of developing CRC provides further evidence that PDE5 is an essential target for CRC chemoprevention [70]. Another consideration is the potential involvement of PDE5 in suppressing tumor immunity, as demonstrated by publications reporting that tadalafil, a PDE5 inhibitor, can activate mechanisms of antitumor immunity [64, 71-73]. Thus, PDE5 may be a promising molecular target for the chemoprevention of CRC, but further investigation is needed to understand better if the mechanism involves a direct effect on neoplastic cells or an indirect effect involving the activation of antitumor immunity.

3.2. Role of PDE10 in regulating CRC cell growth

PDE10 is a dual cAMP and cGMP degrading PDE isozyme that has limited expression in peripheral tissues compared with other PDE isozymes [74-76], but is highly expressed in the brain and localized in the medium spiny neurons of the striatum [76, 77]. PDE10 is known to regulate glutamatergic stimulation through the striatal pathway [78]; however, there is no known physiological role of PDE10 outside of the central nervous system. Preclinical studies showed that PDE10 inhibitors may affect cognition, behavior, and motor functions, as demonstrated in rodent models and genetic knockout experiments [79], and were developed for treating schizophrenia and Huntington's disease. PDE10 also has distinct kinetic properties from other PDE isozymes with higher affinity but lower Vmax for cAMP than cGMP [80].

Despite low expression in most normal peripheral tissues, PDE10 was reported to be overexpressed in colon adenomas, adenocarcinomas, and metastatic lesions compared to normal colonic mucosa [81-83]. An essential role of PDE10 in CRC cell growth was first reported by Li and colleagues [81]. For example, known highly specific PDE10 inhibitors were found to selectively inhibit the proliferation and induce apoptosis of CRC cells with reduced effects on the growth of normal colonocytes, which correlated with differences in PDE10 expression. Consistent with the effect of PDE10 inhibitors, PDE10 siRNA also selectively inhibited growth and induced apoptosis of CRC cells with reduced effects on normal colonocytes. Conversely, ectopic expression of PDE10 in normal colonocytes or cells derived from colon adenomas increased the growth rate [81]. PDE10 inhibitors and siRNA knockdown of PDE10 also increased intracellular cGMP levels, activated PKG and suppressed β -catenin transcriptional activity in a manner as described above for PDE5 inhibitors.

A novel non-COX inhibitory derivative of sulindac with PDE10 isozyme selectivity, ADT-061, was developed that potently inhibits the growth of CRC cells expressing PDE10, but not the growth of normal colonocytes lacking PDE10 expression. PDE10 inhibition by ADT-061 was associated with the activation of cGMP/PKG signaling and the phosphorylation and degradation of β -catenin, thereby reducing nuclear levels of β -catenin and TCF/LEF transcriptional activity [84]. In addition to suppressing β -catenin transcriptional activity, PDE10 inhibitors have been reported to activate dendritic cells and increase T-cell tumor invasion, suggesting the potential to generate anticancer immune responses [85, 86]. ADT-061 (aka MCI-030) was also reported to inhibit the growth of ovarian cancer cells by suppressing Wnt/ β -catenin and RAS/MAPK signaling [87], both of which may result from its PDE10 inhibitory activity as previously reported for known PDE10 inhibitors [82], possibly by a cGMP/PKG dependent mechanism to block RAS activation [88]. Alternatively, the inhibitory effect on RAS/MAPK signaling may result from a distinct effect on RAS, as suggested by previous studies of sulindac [19, 26].

3.3. Non-COX-inhibitory sulindac derivatives that inhibit both PDE5 and PDE10

Since CRC cells co-express PDE5 and PDE10 to regulate intracellular cGMP levels, dual PDE5/10 inhibitors would be expected to have more significant growth inhibitory activity than selective inhibitors of PDE5 or PDE10, given the possibility that the uninhibited isozyme remains active to diminish sensitivity to an isozyme specific inhibitor. The co-expression of PDE5 and PDE10 in CRC cells may also explain why high doses of the PDE5 selective inhibitor, sildenafil, are required to inhibit CRC cell growth *in vitro* and tumor growth in a subcutaneous mouse cancer model [66]. For example, uninhibited PDE10 could rapidly degrade cGMP that results from a PDE5 specific inhibitor, necessitating high dosages to non-selectively inhibit PDE10. Conversely, PDE5 co-expression with PDE10 may contribute to resistance to PDE10 selective inhibitors (e.g., Pf2545920), as uninhibited PDE5 could rapidly degrade cGMP generated by a particular PDE10 inhibitor. Supporting the possibility that a dual inhibitor might have advantages, we reported that genetic knockdown of both PDE5 and PDE10 in CRC cells or treatment with a dual PDE5/PDE10 inhibitor, ADT-094, resulted in more significant growth inhibitory activity compared with single

knockdown of PDE5 or PDE10 or selective PDE5 or PDE10 inhibitors [89]. The mechanism by which ADT-094 inhibits CRC cell growth involves binding to both PDE5 and PDE10 to activate cGMP/PKG signaling, which suppresses Wnt-induced β -catenin/Tcf-mediated transcriptional activity by phosphorylation of amino acid residues known to, induce ubiquitination and proteasomal degradation of β -catenin.

4. Conclusion

Multiple lines of evidence from preclinical, epidemiological, and clinical trials report that NSAIDs have robust antineoplastic activity, but their long-term use for cancer chemoprevention is not approved because of potentially fatal toxicities resulting from COX inhibition and suppression of physiological prostaglandins. While numerous investigators have concluded that the antineoplastic activity of sulindac and other NSAIDs is unrelated to their COX inhibitory activity, the potential to develop safer and more efficacious derivatives for CRC chemoprevention rests on identifying the underlying mechanism of action. This review focused on non-COX inhibitory sulindac derivatives that can selectively suppress the growth of CRC cells by inhibiting cGMP PDE isozymes, PDE5 or PDE10, resulting in the activation of cGMP/PKG signaling and suppression of Wnt/ β -catenin transcriptional activity, to block the expression of proteins essential for the proliferation and survival of cancer cells.

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Conflict of interest

All the authors claim that the manuscript is entirely original. Drs. Chen, Keeton, and Piazza are co-founders and co-owners of ADT Pharmaceuticals LLC. All other authors declare no conflict of interest.

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References

1. Martínez ME, McPherson RS, Levin B, Annegers JF: Aspirin and other nonsteroidal anti-inflammatory drugs and risk of colorectal adenomatous polyps among endoscoped individuals. *Cancer Epidemiol Biomarkers Prev* 1995, 4(7):703-707. <https://pubmed.ncbi.nlm.nih.gov/8672985/>

2. Rosenberg L, Louik C, Shapiro S: Nonsteroidal antiinflammatory drug use and reduced risk of large bowel carcinoma. *Cancer* 1998, 82(12):2326-2333. [https://doi.org/10.1002/\(SICI\)1097-0142\(19980615\)82:12%3C2326::AID-CNCR5%3E3.0.CO;2-Q](https://doi.org/10.1002/(SICI)1097-0142(19980615)82:12%3C2326::AID-CNCR5%3E3.0.CO;2-Q)
3. Muscat JE, Stellman SD, Wynder EL: Nonsteroidal antiinflammatory drugs and colorectal cancer. *Cancer* 1994, 74(7):1847-1854. [https://doi.org/10.1002/1097-0142\(19941001\)74:7%3C1847::AID-CNCR2820740704%3E3.0.CO;2-%23](https://doi.org/10.1002/1097-0142(19941001)74:7%3C1847::AID-CNCR2820740704%3E3.0.CO;2-%23)
4. Collet JP, Sharpe C, Belzile E, Boivin JF, Hanley J, Abenhaim L: Colorectal cancer prevention by non-steroidal anti-inflammatory drugs: effects of dosage and timing. *Br J Cancer* 1999, 81(1):62-68. <https://doi.org/10.1038/sj.bjc.6690651>
5. Isomäki HA, Hakulinen T, Joutsenlahti U: Excess risk of lymphomas, leukemia and myeloma in patients with rheumatoid arthritis. *J Chronic Dis* 1978, 31(11):691-696. [https://doi.org/10.1016/0021-9681\(78\)90071-1](https://doi.org/10.1016/0021-9681(78)90071-1)
6. Friedman GD, Coates AO, Potter JD, Slattery ML: Drugs and colon cancer. *Pharmacoepidemiol Drug Saf* 1998, 7(2):99-106. [https://doi.org/10.1002/\(SICI\)1099-1557\(199803/04\)7:2%3C99::AID-PDS320%3E3.0.CO;2-0](https://doi.org/10.1002/(SICI)1099-1557(199803/04)7:2%3C99::AID-PDS320%3E3.0.CO;2-0)
7. Labayle D, Fischer D, Vielh P, Drouhin F, Pariente A, Bories C, Duhamel O, Troussset M, Attali P: Sulindac causes regression of rectal polyps in familial adenomatous polyposis. *Gastroenterology* 1991, 101(3):635-639. [https://doi.org/10.1016/0016-5085\(91\)90519-Q](https://doi.org/10.1016/0016-5085(91)90519-Q)
8. Giardiello FM, Hamilton SR, Krush AJ, Piantadosi S, Hyland LM, Celano P, Booker SV, Robinson CR, Offerhaus GJ: Treatment of colonic and rectal adenomas with sulindac in familial adenomatous polyposis. *N Engl J Med* 1993, 328(18):1313-1316. <https://doi.org/10.1056/nejm199305063281805>
9. Gurpinar E, Grizzle WE, Piazza GA: NSAIDs inhibit tumorigenesis, but how? *Clin Cancer Res* 2014, 20(5):1104-1113. <https://doi.org/10.1158/1078-0432.Ccr-13-1573>
10. Piazza GA, Alberts DS, Hixson LJ, Paranka NS, Li H, Finn T, Bogert C, Guillen JM, Brendel K, Gross PH et al: Sulindac Sulfone Inhibits Azoxymethane-induced Colon Carcinogenesis in Rats without Reducing Prostaglandin Levels. *Cancer Research* 1997, 57(14):2909-2915. <https://pubmed.ncbi.nlm.nih.gov/9230200/>
11. Nugent KP, Farmer KC, Spigelman AD, Williams CB, Phillips RK: Randomized controlled trial of the effect of sulindac on duodenal and rectal polyposis and cell proliferation in patients with familial adenomatous polyposis. *Br J Surg* 1993, 80(12):1618-1619. <https://doi.org/10.1002/bjs.1800801244>
12. Steinbach G, Lynch PM, Phillips RK, Wallace MH, Hawk E, Gordon GB, Wakabayashi N, Saunders B, Shen Y, Fujimura T et al: The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N Engl J Med* 2000, 342(26):1946-1952. <https://doi.org/10.1056/nejm200006293422603>
13. Tsujii M, DuBois RN: Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell* 1995, 83(3):493-501. [https://doi.org/10.1016/0092-8674\(95\)90127-2](https://doi.org/10.1016/0092-8674(95)90127-2)
14. Tsujii M, Kawano S, Tsuji S, Sawaoka H, Hori M, DuBois RN: Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 1998, 93(5):705-716. [https://doi.org/10.1016/s0092-8674\(00\)81433-6](https://doi.org/10.1016/s0092-8674(00)81433-6)
15. Giardiello FM, Offerhaus GJ, DuBois RN: The role of nonsteroidal anti-inflammatory drugs in colorectal cancer prevention. *Eur J Cancer* 1995, 31a(7-8):1071-1076. [https://doi.org/10.1016/0959-8049\(95\)00137-8](https://doi.org/10.1016/0959-8049(95)00137-8)
16. Oshima M, Dinchuk JE, Kargman SL, Oshima H, Hancock B, Kwong E, Trzaskos JM, Evans JF, Taketo MM: Suppression of intestinal polyposis in Apc delta716 knockout mice by inhibition of cyclooxygenase 2 (COX-2). *Cell* 1996, 87(5):803-809. [https://doi.org/10.1016/s0092-8674\(00\)81988-1](https://doi.org/10.1016/s0092-8674(00)81988-1)
17. Reddy BS, Kawamori T, Lubet RA, Steele VE, Kelloff GJ, Rao CV: Chemopreventive efficacy of sulindac sulfone against colon cancer depends on time of administration during carcinogenic process. *Cancer Res* 1999, 59(14):3387-3391. <https://pubmed.ncbi.nlm.nih.gov/10416599/>
18. Wargovich MJ, Jimenez A, McKee K, Steele VE, Velasco M, Woods J, Price R, Gray K, Kelloff GJ: Efficacy of potential chemopreventive agents on rat colon aberrant crypt formation and progression. *Carcinogenesis* 2000, 21(6):1149-1155. <https://doi.org/10.1093/carcin/21.5.149>
19. Thompson HJ, Jiang C, Lu J, Mehta RG, Piazza GA, Paranka NS, Pamukcu R, Ahnen DJ: Sulfone metabolite of sulindac inhibits mammary carcinogenesis. *Cancer Res* 1997, 57(2):267-271. <https://pubmed.ncbi.nlm.nih.gov/9000566/>
20. Malkinson AM, Koski KM, Dwyer-Nield LD, Rice PL, Rioux N, Castonguay A, Ahnen DJ, Thompson H, Pamukcu R, Piazza GA: Inhibition of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-induced mouse lung tumor formation by FGN-1 (sulindac sulfone). *Carcinogenesis* 1998, 19(8):1353-1356. <https://doi.org/10.1093/carcin/19.8.1353>

21. Piazza GA, Thompson WJ, Pamukcu R, Alila HW, Whitehead CM, Liu L, Fetter JR, Gresh WE, Jr., Klein-Szanto AJ, Farnell DR et al: Exisulind, a Novel Proapoptotic Drug, Inhibits Rat Urinary Bladder Tumorigenesis1. *Cancer Research* 2001, 61(10):3961-3968. <https://pubmed.ncbi.nlm.nih.gov/11358813/>
22. Narayanan BA, Reddy BS, Bosland MC, Nargi D, Horton L, Randolph C, Narayanan NK: Exisulind in combination with celecoxib modulates epidermal growth factor receptor, cyclooxygenase-2, and cyclin D1 against prostate carcinogenesis: in vivo evidence. *Clin Cancer Res* 2007, 13(19):5965-5973. <https://doi.org/10.1158/1078-0432.Ccr-07-0744>
23. McEntee MF, Chiu CH, Whelan J: Relationship of beta-catenin and Bcl-2 expression to sulindac-induced regression of intestinal tumors in Min mice. *Carcinogenesis* 1999, 20(4):635-640. <https://doi.org/10.1093/carcin/20.4.635>
24. He TC, Chan TA, Vogelstein B, Kinzler KW: PPARdelta is an APC-regulated target of nonsteroidal anti-inflammatory drugs. *Cell* 1999, 99(3):335-345. [https://doi.org/10.1016/s0092-8674\(00\)81664-5](https://doi.org/10.1016/s0092-8674(00)81664-5)
25. Baek SJ, Kim KS, Nixon JB, Wilson LC, Eling TE: Cyclooxygenase inhibitors regulate the expression of a TGF-beta superfamily member that has proapoptotic and antitumorigenic activities. *Mol Pharmacol* 2001, 59(4):901-908. <https://doi.org/10.1124/MOL.59.4.901>
26. Herrmann C, Block C, Geisen C, Haas K, Weber C, Winde G, Möröy T, Müller O: Sulindac sulfide inhibits Ras signaling. *Oncogene* 1998, 17(14):1769-1776. <https://doi.org/10.1038/sj.onc.1202085>
27. Stark LA, Reid K, Sansom OJ, Din FV, Guichard S, Mayer I, Jodrell DI, Clarke AR, Dunlop MG: Aspirin activates the NF-kappaB signalling pathway and induces apoptosis in intestinal neoplasia in two in vivo models of human colorectal cancer. *Carcinogenesis* 2007, 28(5):968-976. <https://doi.org/10.1093/carcin/bgl220>
28. Boon EMJ, Keller JJ, Wormhoudt TAM, Giardiello FM, Offerhaus GJA, van der Neut R, Pals ST: Sulindac targets nuclear beta-catenin accumulation and Wnt signalling in adenomas of patients with familial adenomatous polyposis and in human colorectal cancer cell lines. *British Journal of Cancer* 2004, 90(1):224-229. <https://doi.org/10.1038/sj.bjc.6601505>
29. Barker N, Clevers H: Mining the Wnt pathway for cancer therapeutics. *Nat Rev Drug Discov* 2006, 5(12):997-1014. <https://doi.org/10.1038/nrd2154>
30. Lawson KR, Ignatenko NA, Piazza GA, Cui H, Gerner EW: Influence of K-ras activation on the survival responses of Caco-2 cells to the chemopreventive agents sulindac and difluoromethylornithine. *Cancer Epidemiol Biomarkers Prev* 2000, 9(11):1155-1162. <https://pubmed.ncbi.nlm.nih.gov/11097222/>
31. Rice PL, Kelloff J, Sullivan H, Driggers LJ, Beard KS, Kuwada S, Piazza G, Ahnen DJ: Sulindac metabolites induce caspase- and proteasome-dependent degradation of beta-catenin protein in human colon cancer cells. *Molecular Cancer Therapeutics* 2003, 2(9):885-892. <https://pubmed.ncbi.nlm.nih.gov/14555707/>
32. Thompson WJ, Piazza GA, Li H, Liu L, Fetter J, Zhu B, Sperl G, Ahnen D, Pamukcu R: Exisulind induction of apoptosis involves guanosine 3',5'-cyclic monophosphate phosphodiesterase inhibition, protein kinase G activation, and attenuated beta-catenin. *Cancer Res* 2000, 60(13):3338-3342. <https://pubmed.ncbi.nlm.nih.gov/10910034/>
33. Yi B, Chang H, Ma R, Feng X, Li W, Piazza GA, Xi Y: Inhibition of breast cancer cell motility with a non-cyclooxygenase inhibitory derivative of sulindac by suppressing TGFbeta/miR-21 signaling. *Oncotarget* 2016, 7(7):7979-7992. <https://doi.org/10.18632/oncotarget.6888>
34. Jasperson KW, Tuohy TM, Neklason DW, Burt RW: Hereditary and familial colon cancer. *Gastroenterology* 2010, 138(6):2044-2058. <https://doi.org/10.1053/j.gastro.2010.01.054>
35. Stamos JL, Weis WI: The beta-catenin destruction complex. *Cold Spring Harb Perspect Biol* 2013, 5(1):a007898. <https://doi.org/10.1101/cshperspect.a007898>
36. Half E, Bercovich D, Rozen P: Familial adenomatous polyposis. *Orphanet J Rare Dis* 2009, 4:22. <https://doi.org/10.1186/1750-1172-4-22>
37. Bafico A, Liu G, Goldin L, Harris V, Aaronson SA: An autocrine mechanism for constitutive Wnt pathway activation in human cancer cells. *Cancer Cell* 2004, 6(5):497-506. <https://doi.org/10.1016/j.ccr.2004.09.032>
38. Smith G, Carey FA, Beattie J, Wilkie MJ, Lightfoot TJ, Coxhead J, Garner RC, Steele RJ, Wolf CR: Mutations in APC, Kirsten-ras, and p53--alternative genetic pathways to colorectal cancer. *Proc Natl Acad Sci USA* 2002, 99(14):9433-9438. <https://doi.org/10.1073/pnas.122612899>
39. Lynch PM, Ayers GD, Hawk E, Richmond E, Eagle C, Woloj M, Church J, Hasson H, Patterson S, Half E et al: The safety and efficacy of celecoxib in children with familial adenomatous polyposis. *Am J Gastroenterol* 2010, 105(6):1437-1443. <https://doi.org/10.1038/ajg.2009.758>
40. Phillips RK, Wallace MH, Lynch PM, Hawk E, Gordon GB, Saunders BP, Wakabayashi N, Shen Y, Zimmerman S, Godio L et al: A randomised, double blind, placebo controlled study of celecoxib, a selective cyclooxygenase 2

- inhibitor, on duodenal polyposis in familial adenomatous polyposis. *Gut* 2002, 50(6):857-860. <https://doi.org/10.1136/gut.50.6.857>
41. Solomon SD, McMurray JJ, Pfeffer MA, Wittes J, Fowler R, Finn P, Anderson WF, Zauber A, Hawk E, Bertagnoli M: Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *N Engl J Med* 2005, 352(11):1071-1080. <https://doi.org/10.1056/NEJMoa050405>
 42. Vane JR, Bakhle YS, Botting RM: Cyclooxygenases 1 and 2. *Annu Rev Pharmacol Toxicol* 1998, 38:97-120. <https://doi.org/10.1146/annurev.pharmtox.38.1.97>
 43. Dubois RN, Abramson SB, Crofford L, Gupta RA, Simon LS, Van De Putte LB, Lipsky PE: Cyclooxygenase in biology and disease. *Faseb j* 1998, 12(12):1063-1073. <https://doi.org/10.1096/fasebj.12.12.1063>
 44. Chinery R, Coffey RJ, Graves-Deal R, Kirkland SC, Sanchez SC, Zackert WE, Oates JA, Morrow JD: Prostaglandin J2 and 15-deoxy-delta12,14-prostaglandin J2 induce proliferation of cyclooxygenase-depleted colorectal cancer cells. *Cancer Res* 1999, 59(11):2739-2746. <https://pubmed.ncbi.nlm.nih.gov/10364000/>
 45. Sheng H, Shao J, Morrow JD, Beauchamp RD, DuBois RN: Modulation of apoptosis and Bcl-2 expression by prostaglandin E2 in human colon cancer cells. *Cancer Res* 1998, 58(2):362-366. <https://pubmed.ncbi.nlm.nih.gov/9443418/>
 46. Masferrer JL, Leahy KM, Koki AT, Zweifel BS, Settle SL, Woerner BM, Edwards DA, Flickinger AG, Moore RJ, Seibert K: Antiangiogenic and antitumor activities of cyclooxygenase-2 inhibitors. *Cancer Res* 2000, 60(5):1306-1311. <https://pubmed.ncbi.nlm.nih.gov/10728691/>
 47. Nussmeier NA, Whelton AA, Brown MT, Langford RM, Hoeft A, Parlow JL, Boyce SW, Verburg KM: Complications of the COX-2 inhibitors parecoxib and valdecoxib after cardiac surgery. *N Engl J Med* 2005, 352(11):1081-1091. <https://doi.org/10.1056/NEJMoa050330>
 48. Fitzgerald GA: Coxibs and cardiovascular disease. *N Engl J Med* 2004, 351(17):1709-1711. <https://doi.org/10.1056/NEJMp048288>
 49. Elder DJ, Halton DE, Hague A, Paraskeva C: Induction of apoptotic cell death in human colorectal carcinoma cell lines by a cyclooxygenase-2 (COX-2)-selective nonsteroidal anti-inflammatory drug: independence from COX-2 protein expression. *Clin Cancer Res* 1997, 3(10):1679-1683. <https://doi.org/10.0000/PMID9815550>
 50. Kusunuma H, Matsuyuki H, Matsuura M, Imayoshi T, Okumoto T, Matsui H: Induction of apoptotic DNA fragmentation by nonsteroidal anti-inflammatory drugs in cultured rat gastric mucosal cells. *Eur J Pharmacol* 1998, 360(2-3):273-280. [https://doi.org/10.1016/s0014-2999\(98\)00679-7](https://doi.org/10.1016/s0014-2999(98)00679-7)
 51. Hanif R, Pittas A, Feng Y, Koutsos MI, Qiao L, Staiano-Coico L, Shiff SI, Rigas B: Effects of nonsteroidal anti-inflammatory drugs on proliferation and on induction of apoptosis in colon cancer cells by a prostaglandin-independent pathway. *Biochem Pharmacol* 1996, 52(2):237-245. [https://doi.org/10.1016/0006-2952\(96\)00181-5](https://doi.org/10.1016/0006-2952(96)00181-5)
 52. de Mello MC, Bayer BM, Beaven MA: Evidence that prostaglandins do not have a role in the cytostatic action of anti-inflammatory drugs. *Biochem Pharmacol* 1980, 29(3):311-318. [https://doi.org/10.1016/0006-2952\(80\)90506-7](https://doi.org/10.1016/0006-2952(80)90506-7)
 53. Tinsley HN, Gary BD, Thaiparambil J, Li N, Lu W, Li Y, Maxuitenko YY, Keeton AB, Piazza GA: Colon tumor cell growth-inhibitory activity of sulindac sulfide and other nonsteroidal anti-inflammatory drugs is associated with phosphodiesterase 5 inhibition. *Cancer Prev Res (Phila)* 2010, 3(10):1303-1313. <https://doi.org/10.1158/1940-6207.Capr-10-0030>
 54. Williams CS, Watson AJ, Sheng H, Helou R, Shao J, DuBois RN: Celecoxib prevents tumor growth in vivo without toxicity to normal gut: lack of correlation between in vitro and in vivo models. *Cancer Res* 2000, 60(21):6045-6051. <https://pubmed.ncbi.nlm.nih.gov/11085526/>
 55. Stoner GD, Budd GT, Ganapathi R, DeYoung B, Kresty LA, Nitert M, Fryer B, Church JM, Provencher K, Pamukcu R et al: Sulindac sulfone induced regression of rectal polyps in patients with familial adenomatous polyposis. *Adv Exp Med Biol* 1999, 470:45-53. https://doi.org/10.1007/978-1-4615-4149-3_5
 56. Arber N, Kuwada S, Leshno M, Sjodahl R, Hultcrantz R, Rex D: Sporadic adenomatous polyp regression with exisulind is effective but toxic: a randomised, double blind, placebo controlled, dose-response study. *Gut* 2006, 55(3):367-373. <https://doi.org/10.1136/gut.2004.061432>
 57. Piazza GA, Rahm AL, Krutzsch M, Sperl G, Paranka NS, Gross PH, Brendel K, Burt RW, Alberts DS, Pamukcu R et al: Antineoplastic drugs sulindac sulfide and sulfone inhibit cell growth by inducing apoptosis. *Cancer Res* 1995, 55(14):3110-3116. <https://pubmed.ncbi.nlm.nih.gov/7606732/>
 58. Sun W, Stevenson JP, Gallo JM, Redlinger M, Haller D, Algazy K, Giantonio B, Alila H, O'Dwyer PJ: Phase I and pharmacokinetic trial of the proapoptotic sulindac analog CP-461 in patients with advanced cancer. *Clin Cancer Res* 2002, 8(10):3100-3104. <https://scholars.mssm.edu/en/publications/phase-i-and-pharmacokinetic-trial-of-the-proapoptotic-sulindac-an-2>

59. Piazza GA, Keeton AB, Tinsley HN, Gary BD, Whitt JD, Mathew B, Thaiparambil J, Coward L, Gorman G, Li Y et al: A novel sulindac derivative that does not inhibit cyclooxygenases but potently inhibits colon tumor cell growth and induces apoptosis with antitumor activity. *Cancer Prev Res (Phila)* 2009, 2(6):572-580. <https://doi.org/10.1158/1940-6207.Capr-09-0001>
60. Zhang Y, Zhang J, Wang L, Quealy E, Gary BD, Reynolds RC, Piazza GA, Lü J: A novel sulindac derivative lacking cyclooxygenase-inhibitory activities suppresses carcinogenesis in the transgenic adenocarcinoma of mouse prostate model. *Cancer Prev Res (Phila)* 2010, 3(7):885-895. <https://doi.org/10.1158/1940-6207.Capr-09-0273>
61. Tinsley HN, Mathew B, Chen X, Maxuitenko YY, Li N, Lowe WM, Whitt JD, Zhang W, Gary BD, Keeton AB et al: Novel Non-Cyclooxygenase Inhibitory Derivative of Sulindac Inhibits Breast Cancer Cell Growth In Vitro and Reduces Mammary Tumorigenesis in Rats. *Cancers* 2023, 15(3):646. <https://www.mdpi.com/2072-6694/15/3/646>
62. Tinsley HN, Gary BD, Keeton AB, Lu W, Li Y, Piazza GA: Inhibition of PDE5 by sulindac sulfide selectively induces apoptosis and attenuates oncogenic Wnt/ β -catenin-mediated transcription in human breast tumor cells. *Cancer Prev Res (Phila)* 2011, 4(8):1275-1284. <https://doi.org/10.1158/1940-6207.Capr-11-0095>
63. Whitt JD, Li N, Tinsley HN, Chen X, Zhang W, Li Y, Gary BD, Keeton AB, Xi Y, Abadi AH et al: A novel sulindac derivative that potently suppresses colon tumor cell growth by inhibiting cGMP phosphodiesterase and β -catenin transcriptional activity. *Cancer Prev Res (Phila)* 2012, 5(6):822-833. <https://doi.org/10.1158/1940-6207.Capr-11-0559>
64. Piazza GA, Ward A, Chen X, Maxuitenko Y, Coley A, Aboeella NS, Buchsbaum DJ, Boyd MR, Keeton AB, Zhou G: PDE5 and PDE10 inhibition activates cGMP/PKG signaling to block Wnt/ β -catenin transcription, cancer cell growth, and tumor immunity. *Drug Discov Today* 2020, 25(8):1521-1527. <https://doi.org/10.1016/j.drudis.2020.06.008>
65. Li N, Xi Y, Tinsley HN, Gurpinar E, Gary BD, Zhu B, Li Y, Chen X, Keeton AB, Abadi AH et al: Sulindac selectively inhibits colon tumor cell growth by activating the cGMP/PKG pathway to suppress Wnt/ β -catenin signaling. *Mol Cancer Ther* 2013, 12(9):1848-1859. <https://doi.org/10.1158/1535-7163.Mct-13-0048>
66. Mei XL, Yang Y, Zhang YJ, Li Y, Zhao JM, Qiu JG, Zhang WJ, Jiang QW, Xue YQ, Zheng DW et al: Sildenafil inhibits the growth of human colorectal cancer in vitro and in vivo. *Am J Cancer Res* 2015, 5(11):3311-3324. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4697679/>
67. Shi Z, Tiwari AK, Shukla S, Robey RW, Singh S, Kim IW, Bates SE, Peng X, Abraham I, Ambudkar SV et al: Sildenafil reverses ABCB1- and ABCG2-mediated chemotherapeutic drug resistance. *Cancer Res* 2011, 71(8):3029-3041. <https://doi.org/10.1158/0008-5472.Can-10-3820>
68. Lin S, Wang J, Wang L, Wen J, Guo Y, Qiao W, Zhou J, Xu G, Zhi F: Phosphodiesterase-5 inhibition suppresses colonic inflammation-induced tumorigenesis via blocking the recruitment of MDSC. *Am J Cancer Res* 2017, 7(1):41-52. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5250679/>
69. Islam BN, Sharman SK, Hou Y, Bridges AE, Singh N, Kim S, Kolhe R, Trillo-Tinoco J, Rodriguez PC, Berger FG et al: Sildenafil Suppresses Inflammation-Driven Colorectal Cancer in Mice. *Cancer Prev Res (Phila)* 2017, 10(7):377-388. <https://doi.org/10.1158/1940-6207.Capr-17-0015>
70. Huang W, Sundquist J, Sundquist K, Ji J: Use of Phosphodiesterase 5 Inhibitors Is Associated With Lower Risk of Colorectal Cancer in Men With Benign Colorectal Neoplasms. *Gastroenterology* 2019, 157(3):672-681.e674. <https://doi.org/10.1053/j.gastro.2019.05.012>
71. Serafini P, Meckel K, Kelso M, Noonan K, Califano J, Koch W, Dolcetti L, Bronte V, Borrello I: Phosphodiesterase-5 inhibition augments endogenous antitumor immunity by reducing myeloid-derived suppressor cell function. *J Exp Med* 2006, 203(12):2691-2702. <https://doi.org/10.1084/jem.20061104>
72. Noonan KA, Ghosh N, Rudraraju L, Bui M, Borrello I: Targeting immune suppression with PDE5 inhibition in end-stage multiple myeloma. *Cancer Immunol Res* 2014, 2(8):725-731. <https://doi.org/10.1158/2326-6066.Cir-13-0213>
73. Weed DT, Vella JL, Reis IM, De la Fuente AC, Gomez C, Sargi Z, Nazarian R, Califano J, Borrello I, Serafini P: Tadalafil reduces myeloid-derived suppressor cells and regulatory T cells and promotes tumor immunity in patients with head and neck squamous cell carcinoma. *Clin Cancer Res* 2015, 21(1):39-48. <https://doi.org/10.1158/1078-0432.Ccr-14-1711>
74. Soderling SH, Bayuga SJ, Beavo JA: Isolation and characterization of a dual-substrate phosphodiesterase gene family: PDE10A. *Proc Natl Acad Sci U S A* 1999, 96(12):7071-7076. <https://doi.org/10.1073/pnas.96.12.7071>
75. Omori K, Kotera J: Overview of PDEs and their regulation. *Circ Res* 2007, 100(3):309-327. <https://doi.org/10.1161/01.RES.0000256354.95791.f1>

76. Fujishige K, Kotera J, Michibata H, Yuasa K, Takebayashi S, Okumura K, Omori K: Cloning and characterization of a novel human phosphodiesterase that hydrolyzes both cAMP and cGMP (PDE10A). *J Biol Chem* 1999, 274(26):18438-18445. <https://doi.org/10.1074/jbc.274.26.18438>
77. Seeger TF, Bartlett B, Coskran TM, Culp JS, James LC, Krull DL, Lanfear J, Ryan AM, Schmidt CJ, Strick CA et al: Immunohistochemical localization of PDE10A in the rat brain. *Brain Res* 2003, 985(2):113-126. [https://doi.org/10.1016/s0006-8993\(03\)02754-9](https://doi.org/10.1016/s0006-8993(03)02754-9)
78. Wagner S, Teodoro R, Deuther-Conrad W, Kranz M, Scheunemann M, Fischer S, Wenzel B, Egerland U, Hoefgen N, Steinbach J et al: Radiosynthesis and biological evaluation of the new PDE10A radioligand [18F]AQ28A. *Journal of Labelled Compounds and Radiopharmaceuticals* 2017, 60(1):36-48. <https://doi.org/10.1002/jlcr.3471>
79. Siuciak JA, Chapin DS, Harms JF, Lebel LA, McCarthy SA, Chambers L, Shrikhande A, Wong S, Menniti FS, Schmidt CJ: Inhibition of the striatum-enriched phosphodiesterase PDE10A: a novel approach to the treatment of psychosis. *Neuropharmacology* 2006, 51(2):386-396. <https://doi.org/10.1016/j.neuropharm.2006.04.013>
80. Jäger R, Russwurm C, Schwede F, Genieser HG, Koesling D, Russwurm M: Activation of PDE10 and PDE11 phosphodiesterases. *J Biol Chem* 2012, 287(2):1210-1219. <https://doi.org/10.1074/jbc.M111.263806>
81. Li N, Lee K, Xi Y, Zhu B, Gary BD, Ramírez-Alcántara V, Gurpinar E, Canzoneri JC, Fajardo A, Sigler S et al: Phosphodiesterase 10A: a novel target for selective inhibition of colon tumor cell growth and β -catenin-dependent TCF transcriptional activity. *Oncogene* 2015, 34(12):1499-1509. <https://doi.org/10.1038/onc.2014.94>
82. Zhu B, Lindsey A, Li N, Lee K, Ramirez-Alcantara V, Canzoneri JC, Fajardo A, Madeira da Silva L, Thomas M, Piazza JT et al: Phosphodiesterase 10A is overexpressed in lung tumor cells and inhibitors selectively suppress growth by blocking β -catenin and MAPK signaling. *Oncotarget* 2017, 8(41):69264-69280. <https://doi.org/10.18632/oncotarget.20566>
83. Lee K, Lindsey AS, Li N, Gary B, Andrews J, Keeton AB, Piazza GA: β -catenin nuclear translocation in colorectal cancer cells is suppressed by PDE10A inhibition, cGMP elevation, and activation of PKG. *Oncotarget* 2016, 7(5):5353-5365. <https://doi.org/10.18632/oncotarget.6705>
84. Lee KJ, Chang WL, Chen X, Valiyaveetil J, Ramirez-Alcantara V, Gavin E, Musiyenko A, Madeira da Silva L, Annamdevula NS, Leavesley SJ et al: Suppression of Colon Tumorigenesis in Mutant Apc Mice by a Novel PDE10 Inhibitor that Reduces Oncogenic β -Catenin. *Cancer Prev Res (Phila)* 2021, 14(11):995-1008. <https://doi.org/10.1158/1940-6207.Capr-21-0208>
85. Spranger S, Bao R, Gajewski TF: Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity. *Nature* 2015, 523(7559):231-235. <https://doi.org/10.1038/nature14404>
86. Spranger S, Dai D, Horton B, Gajewski TF: Tumor-Residing Batf3 Dendritic Cells Are Required for Effector T Cell Trafficking and Adoptive T Cell Therapy. *Cancer Cell* 2017, 31(5):711-723.e714. <https://doi.org/10.1016/j.ccell.2017.04.003>
87. Borneman RM, Gavin E, Musiyenko A, Richter W, Lee KJ, Crossman DK, Andrews JF, Wilhite AM, McClellan S, Aragon I et al: Phosphodiesterase 10A (PDE10A) as a novel target to suppress β -catenin and RAS signaling in epithelial ovarian cancer. *Journal of Ovarian Research* 2022, 15(1):120. <https://doi.org/10.1186/s13048-022-01050-9>
88. Cho K-j, van der Hoeven D, Zhou Y, Maekawa M, Ma X, Chen W, Fairn GD, Hancock JF: Inhibition of Acid Sphingomyelinase Depletes Cellular Phosphatidylserine and Mislocalizes K-Ras from the Plasma Membrane. *Molecular and Cellular Biology* 2016, 36(2):363-374. <https://doi.org/10.1128/MCB.00719-15>
89. Li N, Chen X, Zhu B, Ramírez-Alcántara V, Canzoneri JC, Lee K, Sigler S, Gary B, Li Y, Zhang W et al: Suppression of β -catenin/TCF transcriptional activity and colon tumor cell growth by dual inhibition of PDE5 and 10. *Oncotarget* 2015, 6(29):27403-27415. <https://doi.org/10.18632/oncotarget.4741>